10.51152/jbarbiomed.v9i1.229



Original Article

Antimicrobial Effects of Sansevieria Zeylanica Extracts on Urinary Tract Infection Associated Pathogens Isolated from Students Attending a Tertiary Institution

Omoruyi Zainab*1, Abdullateef Ibrahim Olalekan1, Egunjobi Tunde Oluwasegun2

¹Department of Medical Laboratory Science, School of Basic Medical Sciences, University of Benin, Benin City, Nigeria.

Received: 15-11-2023 Revised: 24-12-2023 Published: 30-12-2023

Keywords:

Antimicrobial effect, Antibiotics, methanol, Sansevieria zeylanica, Uropathogens

Abstract: Urinary tract infections usually develop in the lower urinary tract and if not properly treated, they ascend to the upper urinary tract, causing severe damage to the kidneys. Sansevieria zeylenica is a plant known to be rich in phenolic compounds, which may be responsible for its antibacterial activities against Uropathogens. The study was aimed to investigate the antimicrobial effect of Sansevieria zeylenica extracts against isolate of UTI among students attending tertiary institution. A total of two hundred (200) urine samples were randomly collected from the students attending University of Benin into sterile universal containers and analyzed for the detection of Uropathogens using standard microbiological procedure while the leaf and root of Sansevieria zeylenica extraction was carried out with methanol using the Soxhlet apparatus. The antimicrobial activity against the isolates was determined using Agar well diffusion and the minimum inhibitory concentration (MIC) of the extracts against the isolates was determined using the microdilution method. Of the 200 participants studied, 23 (11.5%) had growth of four bacteria species associated with urinary tract infection; Escherichia coli, Proteus vulgaris, Pseudomonas aeruginosa, and Staphylococcus aureus. The activity of both leaf and root extracts was greater at 200mg/ml against E. coli and P. vulgaris isolates, but at 400mg/ml against S. aureus with Minimum Inhibitory Concentrations of the leaf extract as 8mg/ml, 4mg/ml, 8mg/ml, and >16mg/ml against E. coli, S. aureus, P. vulgaris, and P. aeruginosa, respectively. The association of leaf extract and Ampicillin increases the activity of Ampicillin, producing a greater zone of inhibition (26mm) against P. vulgaris compared to Ampicillin alone (21mm) and leaf extract alone (16mm) giving rise to a synergistic effect. Sansevieria zeylanica extracts in this study can control Urinary Tract Infection and may be combined with conventional antibiotics as a treatment for Uropathogens. Though, further investigation would be requiring for the molecular mechanism behind its antibacterial activity.

Cite this article as: Zainab, O., Olalekan, A.I. Oluwasegun, E.T. (2023) Antimicrobial Effects of Sansevieria Zeylanica Extracts on Urinary Tract Infection Associated Pathogens Isolated from Students Attending a Tertiary Institution. Journal of Basic and Applied Research in Biomedicine, 9(1): 26-29 10.51152/jbarbiomed.v9i1.229



This work is licensed under a Creative Commons Attribution 4.0 License. You are free to copy, distribute and perform the work. You must attribute the work in the manner specified by the author or licensor.

INTRODUCTION

The plant Sanseveira zeylanica, commonly known as bowstring hemp is a species of plant belonging to the genus sanseveira and family Asparagus (Asparagacear) (Evans 2005). There are more Sansevierians in Africa than anywhere in the world (Carlquist and Schneider, 2007). Many studies have shown that medicinal plants are sources of nutrient and non-nutrient compounds, many of which show antioxidant and antimicrobial properties, which are known to protect the human body against both cellular oxidation reactions and pathogens (Nascimento et al., 2000). In Nigeria, the leaves and roots of Sansevieria species are used traditionally to treat diseases such as diarrhea, eczema, gonorrhea, hemorrhoids and piles (Adeyemi et al., 2009). Sanseveira species are known to exhibit anti-inflammatory, antioxidant, and antimicrobial properties (Aliero et al., 2008), the leaf and rhizome extracts of this plant have the ability to inhibit the growth of many disease causing bacteria and this ability serves a potential of broad spectrum antimicrobial effect which makes the plant suitable for producing antimicrobial drugs (Igbinosa and Aiyegoro, 2009).

Urinary tract infection (UTI) affects close to 150 million people worldwide (Sihra *et al*, 2018). Recurrent UTI (rUTI) affects mostly young female, but it has been estimated that up to 50% of women experience UTI at least once in their lives (Silverman *et al.*, 2013). Furthermore, UTI is mainly associated with patients that have history of urinary tract abnormalities, suppressed immune systems, long-term catheter use, and recent urinary procedures. Several recent studies propound a genetic

susceptibility to the recurrent symptomatic UTI. Numerous genes seem to contribute and have been strongly associated with UTI-prone patients (Murray *et al.*, 2021).

Medicinal plant serves an alternative that has been an effective complementary practical approach for treating rUTI, especially as a prophylactic therapy of antibiotics, as both have synergistic effect (Belkaid and Hand, 2014). Medicinal plant is a realistic option and a better choice for the long-term prevention of rUTI as they are cheap, readily available, safe to use, with fewer reported side effects, and do not cause bacterial resistance (Belkaid and Hand, 2014). The antimicrobial effects of plant extracts involves multiple mechanisms which include directly killing microbes, interfering with microbial adhesion to epithelial cells and biofilm formation, inhibiting the immunomodulators, or boosting body oxidant status (Belkaid, and Hand, 2014). Against this background, this study was conducted to determine the antimicrobial effects of methanolic extracts of sansevieria zeylanica against UTI isolates from tertiary institution.

MATERIALS AND METHODS

Study Design

A cross-sectional study was conducted among students attending University of Benin, Edo state.

Study Area

This study was conducted in Ugbowo campus, University of Benin, Benin city Edo state. The University of Benin, Ugbowo

²Department of Medical Microbiology, School of Basic Clinical Sciences, College of Health Sciences, Igbinedion University Okada, Edo State, Nigeria.

^{*}Corresponding Author: zainab.omoruyi@uniben.edu

campus is in Egor Local Government Area, with a population estimated at 258,442 inhabitants and lies between longitudes 5⁰ 34E and latitude 6⁰ 23N (Ministry of Land and Survey, 2008). The main occupations of the people in these local government areas include trading, farming and private transport system.

Study Population

A total of two hundred (200) students residing in the Ugbowo campus of the University of Benin, Edo State, Nigeria, were recruited for this study. Each student gave written consent for their voluntary participation in the study. Prior to the conduction of the study, ethical approval was obtained from the University of Benin Ethical Committee with the reference number: CMS/REC/2022/305

Urine Sample Collection

Prior to collection of samples, guidelines on proper collection of urine sample to prevent contamination were given to each student and a voided midstream (clean-catch) urine sample was collected from each student into a sterile screw-capped plastic universal container.

Microbiological Analysis

A standardized loop was used to inoculate 0.001ml of properly mixed uncentrifuge urine specimen onto the surface of cysteine lactose electrolyte deficient (CLED) medium and blood agar. The agar plates were incubated aerobically at 37 $^{\circ}$ C, and colonies were counted after 24hours. Urinary tract infection was indicated at colony count $\geq 10^{5}$ colony forming unit/ml, and isolates identified by standard microbiological procedure (Barrow and Feltham, 2003).

Plant Sample Collection

The leaves and roots of *Sansevieria zeylanica* were collected from the garden opposite the laboratory complex of the School of Clinical Medicine, Igbinedion University, Okada (IUO), Edo State and confirmed at the Department of Biological Sciences, IUO.

Preparation of Extracts

The leaves and roots of *Sansevieria zeylanica* were allowed to dry. After drying, 132.23g and 54.81g of the leaves and roots, respectively of *S. zeylanica* were pulverized and the methanolic extraction was carried out with the aid of the Soxhlet apparatus. The extracts were filtered with Whatman No 1 filter paper, concentrated to dryness at 40°C, and stored at 4°C.

Antimicrobial Activity Test

Agar well diffusion method described by White and Reeves (1987) was used to determine the susceptibility of the bacterial isolates to the plant extracts. The minimum inhibitory concentration (MIC) of the extracts against the isolates was determined using the microdilution method as previously described (Eloff, 1998). Varying concentrations of the extracts ranging from 0.125mg/ml to 16mg/ml were prepared in Muller-Hinton broth medium. A 0.1ml of the standardized suspension of each bacterial isolate was inoculated into each tube and incubated aerobically at 37°C for 24 hours. The tube with the lowest concentration without visible growth, when compared with the control (tube containing broth medium and extract only), was regarded as the minimum inhibitory concentration (MIC).

Extract and Antibiotic Combination Synergism Test

The isolates were subjected to susceptibility test by using selected antibiotic discs impregnated with the plant extracts. The zone of inhibition was measured after 24 hours of incubation and compared with diameters produced by antibiotic alone and extract alone. A synergistic effect was indicated when the zone of inhibition produced by the antibiotic-extract combination was greater than the diameter produced by antibiotic alone or extracts alone, as described by Kingsley *et al* (2013).

RESULTS

Prevalence of Urinary Tract Infection

A total of 200 urine samples were collected out of which only 23(11.5%) yielded the growth of uropathogens (Figure 1). Four bacteria species associated with urinary tract infection were identified; *Escherichia coli*, *Proteus vulgaris*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus*. *Escherichia coli* was the predominant bacteria specie in the study with 43.5% (Figure 2).

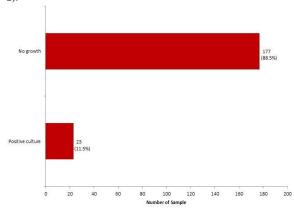


Figure 1: Prevalence of Urinary Tract Infection among the Study Population.

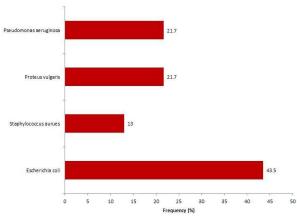


Figure 2: Frequency of pathogens isolated from participant samples.

Antimicrobial Activity of Methanolic Leaf and Root Extracts of Sansevieria zeylanica

The antimicrobial activity of the methanolic extracts of the leaf and root of *S. zeylanica* was tested against four bacterial isolated in this study using the agar well diffusion method. Both the leaf and root extracts showed varying degrees of antimicrobial activity at concentration between 100mg/ml and 400mg/ml (zone of inhibition of 0mm to 20mm) (Table 1). The activity of both extracts was greater at 200mg/ml against *E. coli* and *P. vulgaris* isolates, but at 400mg/ml against *S. aureus*.

The Minimum Inhibitory Concentrations (MICs) of the leaf extract were 8mg/ml, 4mg/ml, 8mg/ml, and >16mg/ml against *E. coli*, *S. aureus*, *P. vulgaris*, and *P. aeruginosa*, respectively (Table 2).

Table 1: Antimicrobial Activity of the Methanolic Extracts of the Leaf and Root of S.

zeylanica with zone of inhibition in millimetre (mm).

Bacteria	Leave Extract Conc.		Root Extract Conc.		AMP	NA		
Isolate	(mg/ml)			(mg/ml)				
	400	200	100	400	200	100		
E. coli	15	15	11	9	13	9	0	0
S. aureus	20	18	9	16	15	10	0	0
P. vulgaris	10	16	9	12	19	10	21	25
Р.	0	0	0	0	0	0	0	0
aeruginosa								

Conc. - Concentration, AMP - Ampicillin, NA - Nalidixic acid.

Table 2: Minimum Inhibitory Concentration (MIC) in mg/mL of the methanolic leaf and root extracts of S. zeylanica.

Bacteria Isolate	Leave	Root
E. coli	8	16
S. aureus	4	8
P. vulgaris	8	8
P. aeruginosa	>16	>16

Synergistic Effect of Antibiotics and Plant Extracts

The combined effect of selected antibiotics and *S. zeylanica* extracts are shown in Tables 3 and 4. Ampicillin and the leaf extract produced a greater zone of inhibition (26mm) against *P. vulgaris* compared to Ampicillin alone (21mm) and leaf extract alone (16mm) indicating a synergistic effect.

Table 3: Combined Effect of the Methanolic Leaf Extract and Selected Antibiotics.

Table 5: Combined Effect of the Methanishe Bear Extract and Selected Intelligence.						
Bacteria	LE	AMP	AMP+LE	NA	NA+LE	
Isolate						
E. coli	15	0	10	0	12	
S. aureus	18	0	13	0	10	
P. vulgaris	16	21	26	25	13	
P.	0	0	0	0	0	
aeruginosa						

LE – 200mg/ml Leave extract, AMP – 10µg/disc Ampicillin, AMP+LE - 10µg/disc Ampicillin and 200mg/ml Leave extract combination, NA – 30µg/disc Nalidixic acid, NA+LE - 30µg/disc Nalidixic acid and 200mg/ml Leave extract combination.

Table 4: Combined Effect of the Methanolic Root Extract and Selected Antibiotics.

Bacteria Isolate	RE	AMP	AMP+RE	NA	NA+RE
E. coli	9	0	7	0	7
S. aureus	15	0	12	0	11
P. vulgaris	19	21	21	25	16
P.	0	0	0	0	0
aeruginosa					

RE – 200mg/ml Root extract, AMP – 10µg/disc Ampicillin, AMP+RE - 10µg/disc Ampicillin and 200mg/ml Root extract combination, NA – 30µg/disc Nalidixic acid, NA+RE - 30µg/disc Nalidixic acid and 200mg/ml Root extract combination

DISCUSSION

Urinary tract infections (UTIs) are the second most common human infections in the body (UDHHS, 2004). With the global spread of Multidrug-resistant (MDR) pathogens, the treatment of UTIs has become difficult resulting in poor prognosis, hence, the need to search for new antimicrobial agents effective against urinary tract pathogens. An overall prevalence of 11.5% of bacteria was observed among participants in this study. This differed from the 49.0% reported by Orrett (2001) in South America but compare with 11.0% reported by Omoregie et al (2008) in Edo state, Nigeria. Geographical location may be the reason for the difference. Escherichia coli is a major pathogen associated with UTI as observed in the present study (43.5%). this is in tandem with a previous study among students of a tertiary institution in Nigeria (Ojo and Anibijuwon, 2010). The high prevalence of E. coli as a causative agent of UTI is often linked to fecal contamination of the urethra, especially in females (Nicolle, 2001).

The methanolic leaf and root extracts of S. zevlanica showed good antimicrobial activity against tested isolates of E. coli, Staphylococcus aureus, and Proteus vulgaris with zone of inhibition ranging from 9mm to 20mm. This finding agrees with other studies that reported the antimicrobial effect of Sansevieria species on these pathogens (Deepa et al., 2011; Tkachenko et al., 2017). Ampicillin and Nalidixic acid are common antibiotics used in the treatment of UTIs, however, it is not uncommon to have uropathogens resistant to these antimicrobial agents (Ojo and Anibijuwon, 2010; Oladehinde et al., 2011). Although isolates of E. coli and S. aureus tested were resistant to Ampicillin and Nalidixic acid, these pathogens were greatly susceptible to the methanolic extracts of S. zeylanica. The resistance of the isolates to these antibiotics could be attributed to the extensive use of these antimicrobial agents over the years. Activity of S. zeylanica and selected antibiotics on Pseudomonas aeruginosa showed resistance to both extracts. In the study of Deepa et al (2011), the methanolic leaf and root extracts of Sansevieria roxburghiana showed antimicrobial activity against P. aeruginosa (zone of inhibition of 12mm). The variations in results could be due to one, differences in the strain of P. aeruginosa tested and site of isolation, and two, the presence or high amount of certain bioactive substance(s) in S. roxburghiana which may be absent or reduced in S. zeylanica. This is consistent with the study by (Ghesmati, 2008) which reported the resistance of strains of P. aeruginosa to certain plant extracts. The emergence of multidrug-resistant (MDR) and extensively drug-resistant Pseudomonas aeruginosa strains associated with human infections particularly UTIs is a public health concern (Breidenstein et al., 2011; Poole, 2011).

The Minimum Inhibitory Concentration (MIC) values of 8mg/ml, 4mg/ml and 8mg/ml inhibited the growth of *Escherichia coli*, *S. aureus* and *P. vulgaris*, respectively with the

methanolic leaf extract. The MIC of the leaf extract against *S. aureus* in the present study corresponds with the report of Ugbomoiko *et al* (2022). Ampicillin and the methanolic leaf extract showed a synergistic effect against *P. vulgaris*. In their study, Kingsley *et al* (2013) reported that Sansevieria species are capable of producing a synergistic effect when combined with certain antibiotics.

In contrast, an antagonistic relationship was observed between Nalidixic acid and extracts of *S. zeylanica* against all the tested bacterial isolates. Darwish *et al* (2002) reported certain plant extracts capable of antagonizing the antibacterial activity of Nalidixic acid. In the same vein, the root extract of *S. zeylanica* has been reported to have an antagonistic effect when combined with Gentamycin (Ugbomoiko *et al.*, 2022). The possible explanation to the antagonistic effect of extracts of *S. zeylanica* on Nalidixic acid may be due to the presence of bioactive substance(s) in the plant inhibitory to the action of the antimicrobial agent. Further studies are needed to verify this point.

CONCLUSION

To the best of our knowledge, this is the first time the antimicrobial effect of extracts of *S. zeylanica* on pathogens associated with UTI is reported. The inhibitory effect of the extracts on both the Gram -positive and -negative bacteria indicates a broad spectrum activity, and the action on the latter is remarkable as these bacteria are major expressers of various resistance genes. Subsequently, the antimicrobial activity of the extracts of *S. zeylanica* against common uropathogens as observed in this study substantiates the traditional use of this plant in the treatment of UTIs.

CONFLICT OF INTEREST

There is no conflict of interest

REFERENCES

Adeyemi, O.O., Akindele, A.J. and Ogunleye, E.A. (2009). Evaluation of the antidiarrhoeal effect of Sanseviera liberica Gerome and Labroy (Agavaceae) root extract. Journal of Ethnopharmacology, 123(3): 459-463.

Aliero, A. A., Jimoh, F and Afolayan, A.J. (2008). Antioxidant and antibacterial properties of Sansevieria hyacinthoides. International Journal of Pure Applied Science. 2:103-110.

Barrow, G. I., Feltham, R. K. A. (2003). Cowan and Steel's manual for the identification of medical bacteria (3rd edition). Cambridge: Cambridge University Press.

Belkaid, Y. and Hand, T.W. (2014). Role of the microbiota in immunity and inflammation. *Cell*, 157, 121–141.

Breidenstein, E. B. M., de la Fuente-Núñez, C., Hancock, R. (2011). Pseudomonas aeruginosa: all roads lead to resistance. Trends in Microbiology. 19:419–426.

Carlquist, S. and Schneider, E.L. (2007). Origins and nature of vessels in monocotyledons. Sansevierias. South Africa Journal of Botany.73:196-203.

Darwish, R. M., Aburjai, T., Al-Khalil, S., Mahafzah, A. (2002). Screening of antibiotic resistant inhibitors from local plant materials against two different strains of *Staphylococcus aureus*. Journal of Ethnopharmacology. 79:359–364

Deepa, P., Kaleena, P.K., Valivittan, K., Kumar, C.P.G. (2011). Phytochemical Screening and Antimicrobial Activity of *Sansevieria roxburghiana* Schult. And Schult. F. Middle East Journal of Scientific Research. 10 (4):512-518.

Eloff, J. N. (1998). Which extract should be used for the screening and isolation of antimicrobial components from plants? Journal of Ethnopharmacology. 60(1): 1-8.

Evans, W.C (2005). Trease and Evans Pharmacognosy, 15th edition. India: Elsevier.

Ghesmati, M. (2008). Survey of antibacterial activity of Sambucusebulus extracts against Staphylococcus aureus ATCC 1341 and Pseudomonas aeruginosa ATCC 2785. Journal of Biological Sciences. 1:73-82.

Igbinosa, O.O, Igbinosa, E.O and Aiyegoro, O.A. (2009).
Antimicrobial activity and phytochemical screening of

- stem bark extracts from Jatropha curcas (Linn). Africa Journal of pharmacy and pharmacology. 3(2):58-62.
- Kingsley, D., Chauhan, R., Sinha, P., Abraham, J. (2013). Screening and Characterization of Antimicrobial Agents from *Sansevieria roxburghiana* and *Sansveria trifasiata*. Asian Journal of Plant Sciences. 12:224-227.
- Murray B.O., Flores, C., Williams C., Flusberg D.A., Marr E.E., Kwiatkowska K.M., Charest J.L, Isenberg B.C and Rohn, J.L. (2021). Recurrent Urinary Tract Infection: A Mystery in Search of Better Model Systems. Frontier Cellular and Infection Microbiology. 11:691210.
- Nascimento, G.F., Locatelli, J., Freitas, P.C. and Silva, G.L. (2000). Antibacterial Activity of Plant Extracts and Phytochemicals on Antibiotic-Resistant Bacteria. Brazillian Journal of Microbiology. 31:1-4.
- Nicolle, L. E. (2001). Epidemiology of urinary tract infection. Journal of Infectious Diseases. 11:551-564.
- Ojo, O. O. and Anibijuwon, I. I. (2010). Urinary tract infection among female students residing in the campus of the University of Ado Ekiti, Nigeria. Africa Journal of Microbiology Research. 4(12):1195-1198.
- Oladeinde, B. H., Omoregie, R., Olley, M., Anunibe, J. A. (2011). Urinary tract infection in a rural community of Nigeria. North America Journal of Medicine and Science. 3:75-77.
- Omoregie, R., Erebor J.O., Ahonkhai I., Isobor J.O., Ogefere, H.O (2008). Observed changes in the prevalence of uropathogens in Benin City, Nigeria. New Zealand Journal of Medical Laboratory Science. 62:29-31.
- Orrett, F.A. (2001). Urinary tract infection in general practice in a rural community in south Trinidad. Saudi Medical Journal. 22(6):537-540.
- Poole, K. (2011). Pseudomonas aeruginosa: resistance to the max. Frontiers in Microbiology. 2:65.
- Sihra, N., Goodman, A., Zakri, R., Sahai, A., Malde, S. (2018). Non antibiotic prevention, and management of recurrent urinary tract infection. Nature Reviews Urology. 15:750– 776.
- Silverman, J.A., Schreiber, H.L., Hooton, T.M and Hultgren, S.J. (2013). From Physiology to Pharmacy: Developments in the Pathogenesis and Treatment of Recurrent Urinary Tract Infection. Current Urology Report. 14:448–456.
- Tkachenko, H., Buyun, L., Osadowski, Z., Maryniuk, M. (2017).

 The Antibacterial Activity of Certain
 SansevieriaThunb.Species against *Escherichia coli*.

 Agrobiodiversity. 446–453.
- UDHHS. (2004). Vital and Health Statistics. 13(1): 157
- Ugbomoiko, D.O., Egunjobi, T.O., Omosigho, P. O., Olley, M. Osaiyuwu, C., Asemota, P.A., Omoruyi, Z. (2022). Antibacterial Activity of the Leaf and Root Extracts of Sansevieria zeylanica Against Strains of Methicillin Sensitive and -Resistant Staphylococcus aureus. International Journal of Current Research in Medical Sciences 8(11): 1 12.
- White, L. O., Reeves, D.S. (1987) Assaying Antibiotics in Pharmaceutical Microbiology, McGraw-Hill Company, New York, 140–145.